Available online at:

http://www.italian-journal-of-mammalogy.it

Research Article

Volume 32 (2): 137-146, 2021



doi:10.4404/hystrix-00452-2021

Population density estimation of meso-mammal carnivores using camera traps without the individual recognition in Maduru Oya National Park, Sri Lanka

Dulan JAYASEKARA^{1,2}, Dharshani MAHAULPATHA^{1,*}, Sriyanie MITHTHAPALA³

¹Department of Zoology, Faculty of Applied Sciences, University of Sri Jayewardenepura, Sri Lanka ²Faculty of Graduate Studies, University of Sri Jayewardenepura, Sri Lanka ³3165/12 Park Road Colombo 5, Sri Lanka

Keywords: Random Encounter Model Camera Trap Distance Sampling population monitoring activity level day range species abundance

Article history: Received: 10 March 2021 Accepted: 23 August 2021

Acknowledgements

We appreciate the generous cooperation of the Maduru Oya National Park staff and the Department of Wildlife Conservation for granting permission to conduct this research. We would also like to express our graittude to the University of Sri Jayewardenepura for the facilities granted and the financial support provided to conduct this research under the university grant ASP/01/RE/SCI/2018/31. We extend our gratitude to Mr. M.R. Mohamed (Former Park Warden, MONP), Mr. Suranga (MONP), team Wildlife Circle and Suweesha Amarakoon for their support on and off the fieldwork. We appreciate the encouraging comments of Dr. U.K.G.K. Padmalal and Prof. Mayuri R. Wijesinghe. We thank Dr. K. Ukuwela and Prof. Akira Mori for their support. We extend special thanks to Dr. Marcus Rowcliffe and Dr. Eric Howe for their comments and insights regarding the REM and CTDS methods. Authors would like to acknowledge the comments, by anonymous reviewers and editors of Hystrix, which helped us to immensely improve upon earlier drafts.

Abstract

Reliable population estimates are crucial for the conservation and management of faunal species. Population data of meso-mammal carnivores in Sri Lanka, as well as elsewhere in the world, is scarce. We estimated population densities of meso-mammal carnivores in Maduru Oya National Park (MONP) using Random Encounter Model (REM) and Camera Trap Distance Sampling (CTDS) methods in this study. A total of 3402 camera trapping days yielded 3357 video captures of 69 different animal taxa including 658 video captures of meso-mammal carnivores. In this study, we recorded all 12 meso-mammal carnivore species found on the island. The two density estimate methods generated similar population estimates indicating that both methods are compatible to be applied in tropical forest habitats for meso-carnivore species. We identify MONP as an area with high richness for the focal species. The study also generated movement speed, activity patterns, activity levels, and day ranges for the focal species, which will be useful for future research. We discuss the population density estimates for different meso-carnivore species and the use of REM and CTDS density estimation methods and their applicability to a tropical meso-carnivore community.

Introduction

Accurate and updated population density estimates are vital for the proper evaluation of the conservation status of species, as well as for the management and decision-making about wildlife populations (Luo et al., 2020; Romairone et al., 2018; Jiménez et al., 2017; Royle et al., 2013; Carbone et al., 2001). Focused research on estimating mammalian carnivore populations remains scarce in Sri Lanka. Although there have been efforts on estimation of the population density of the Sri Lankan leopard (Panthera pardus kotiya) - the apex predator of the country (Webb et al., 2020; Kittle and Watson, 2018; Kittle et al., 2017) — the population densities of many other species of mammalian carnivores have not been assessed (Kittle and Watson, 2018; Miththapala, 2018; Wijesinghe, 2006; Weerakoon and Goonatilake, 2006). In this study, we focused our work on estimating the population densities of meso-mammals of the order Carnivora (meso-carnivores/small carnivores) that inhabit Maduru Oya National Park in the dry zone of Sri Lanka. Meso-mammals are defined as "medium sized mammals larger than rodents, up to roughly fox/jackal sized" (Parker et al., 2012;

Hystrix, the Italian Journal of Mammalogy ISSN 1825-5272 ©⊙⊕©2021 Associazione Teriologica Italiana doi:10.4404/hystrix-00452-2021 Hoffmann et al., 2010), "which are between 150 g–10 kg in weight" (Morrison, 2013).

Several factors such as the difficulty of individual recognition (Johansson et al., 2020) for spatial capture recapture (SCR) density estimate models, nocturnal/elusive behaviour, solitary activity and high costs of live-trapping methods (Hardouin et al., 2021; Romairone et al., 2018; Sheftel, 2018; O'Brien, 2011; Rowcliffe et al., 2008; Silveira et al., 2003) have influenced the lack of information for these species. Meso-mammals of the order Carnivora include an ecologically important guild of species that plays key roles as predators, seed dispersers, as well as influencers of community structures in tropical forest ecosystems, regulating lower trophic levels and maintaining biodiversity (Hardouin et al., 2021; Kittle and Watson, 2018; Kalle et al., 2013; Roemer et al., 2009). They are also considered carriers of diseases, agricultural pests and apex predators in some ecosystems (Roemer et al., 2009). This group of mammals is represented in Sri Lanka by the families Felidae (small wild cats), Herpestidae (mongooses), Viverridae (civets), Mustelidae (otter) and Canidae (jackal). Within these families, there are 12 species (Tab. S1) in Sri Lanka (Hunter, 2019; MoMD&E, 2019; Dittus, 2017; Weerakoon, 2012).

With advancements in camera trapping technology, there has been a rise in research based on camera trapping methods (Green et al., 2020; Meek et al., 2020; Glover-Kapfer et al., 2019; Meek et al., 2014; O'Brien, 2011). The scope of these studies spreads across a wide

DM, DJ and SM conceived the study. DJ and DM devised the methodology and established the camera stations. DJ analyzed the data with input from DM and SM. DJ led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

^{*}Corresponding author

Email address: mahaulpatha@sjp.ac.lk (Dharshani MAHAULPATHA)

range of different ecological facets such as faunal checklists, abundance, density estimations, population monitoring, behavioural studies, species specific focal research studies and wildlife management (Cappelle et al., 2021; Rovero et al., 2013; Meek et al., 2012; Bater et al., 2011; Zimmermann et al., 2011; O'Brien, 2011; TEAM Network, 2011; Clevenger et al., 2009; Tobler et al., 2009; Bowkett et al., 2008; Rovero and De Luca, 2007; Karanth et al., 2006; Sanderson and Trolle, 2005). However, there remained the absence of a reliable and costeffective method of population density estimation of mammalian fauna that cannot be recognised individually (Chatterjee et al., 2020; Gilbert et al., 2020; Rowcliffe et al., 2008; Srbek-Araujo and Chiarello, 2005). This lacuna was filled by the Random Encounter Model (REM) developed by Rowcliffe et al. (2008) after the early efforts of occupancybased models (Royle and Nichols, 2003) and N-mixture models (Royle, 2004) for abundance estimation. Since then, there has been several research studies that have been conducted based on REM model (Palencia et al., 2021b; Pfeffer et al., 2018; Rademaker et al., 2016; Manzo et al., 2012) as well as modified methods such as the Random Encounter and Staying Time (REST) by Nakashima et al. (2018). Spatial count (SC) models (Chandler and Royle, 2013), time-lapse based models (Moeller et al., 2018), spatial presence-absence (SPA) models (Chatterjee et al., 2020; Ramsey et al., 2015) and species space use (SPU) models (Luo et al., 2020) for populations without markings are several other methods that were recently developed each with their own or common limitations. With the rapid technological development of digital camera traps, the video recording capability of camera traps and multiple snapshots with faster trigger speeds have paved the way for development of REST model (Nakashima et al., 2018) and recently, the modified camera trap distance sampling (CTDS) method (Howe et al., 2017) of the well-known "Distance Sampling" (DS) approach (Thomas et al., 2010; Buckland et al., 2015, 2004, 2001).

Instead of using the auxiliary data such as day range determined by telemetry methods to support the REM, during the last decade, this method has evolved to be self-supplemented based solely on camera trapping information (Hofmeester et al., 2017; Rowcliffe et al., 2016, 2011, 2008). The process of calculating the species densities using REM generates several important parameters such as animal speed,

activity level and day range, which then supports a variety of ecological studies. Therefore, REM has provided a means to investigate a wider range of ecological parameters to assist in the species conservation and management.

After the modifications of Howe et al. (2017), the DS method which has been well established over the years - can also be used to determine species densities even when individual markings are absent. Distance sampling can be considered one of the most applied methods for monitoring of wildlife populations (Buckland et al., 2015, 2001; Thomas et al., 2010). However, the traditional DS method was more applicable for species that could be detected easily and directly during the surveys (Corlatti et al., 2020; Buckland et al., 2015). When it comes to rare, elusive and smaller animal species, the applicability was low (Corlatti et al., 2020; Marques et al., 2013). As a result, in the recent past, there has been a rise in usage of passive DS methods such as sonar, radar and acoustic surveys (Corlatti et al., 2020; Buckland et al., 2015; Marques et al., 2013). The implementation of CTDS (another passive DS method) can be considered a revolution in the wildlife population monitoring study methods, as it greatly reduces the limitations that previously prevailed. Availability of user-friendly software and R packages together with adequate methodologies and literature will make CTDS more popular in future camera trap based research work. Since its introduction, CTDS method has generated reliable density estimates in most of the recent studies (Cappelle et al., 2021; Palencia et al., 2021b; Bessone et al., 2020; Harris et al., 2020; Cappelle et al., 2019).

In this study, the SCR methods where individual recognition is required were not selected, because there were no identifiable pelage patterns in most of the focal species except for the Felids. Therefore, as the best alternatives, we selected REM and CTDS methods of density estimation using camera traps. Most of the recent REM and CTDS camera trapping applications have focused on larger ungulate species (Pal et al., 2021; Pfeffer et al., 2018; Rovero and Marshall, 2009) or on single species (Corlatti et al., 2020; Harris et al., 2020; Cappelle et al., 2019; Gray, 2018; Cusack et al., 2015; Anile et al., 2014; Engeman et al., 2013; Manzo et al., 2012). Rich et al. (2019) investigated population density of multiple forest carnivore species, using SCR methods.

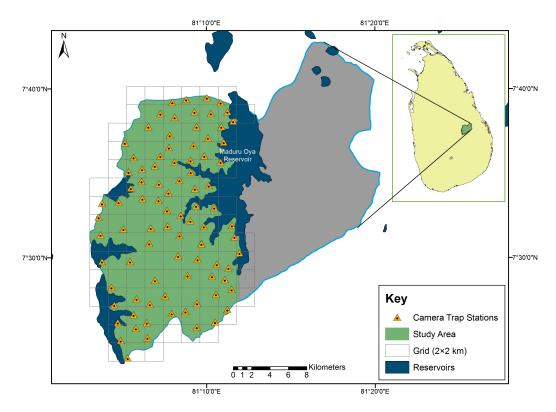


Figure 1 - Map of Maduru Oya National Park with the study area and camera station locations. Location of the Park in the map of Sri Lanka is also shown.

The number of camera trap studies on population densities of mesomammal carnivores remains low and CTDS based multi-species evaluations of this group of fauna are limited (Cappelle et al., 2021; Palencia et al., 2021b; Hardouin et al., 2021; Bessone et al., 2020). Therefore, this is one of the early applications of these new methods to a tropical meso-carnivore community and the first multi-species density estimation in Sri Lanka.

The objectives of this study were; i) to generate density estimates for the meso-mammal carnivores in MONP; ii) to compare the density estimates derived from REM and CTDS methods and assess their applicability in practical situations. During the process of generating density estimates, we developed activity levels, activity patterns, day range, and detection radius/distance parameters for the focal species. Hence, the results generated through this study will provide a range of information to fill research gaps and to benefit future conservation and management requirements.

Materials and Methods

Study area

We conducted this study in Maduru Oya National Park (588 km²) situated in the dry zone (predominantly, in the northern and eastern parts of the country) (Punyawardena, 2020) of Sri Lanka. We carried out camera trapping in the western flank of the park adjacent to the western bank of the Maduru Oya reservoir situated in the centre of the park (Fig. 1). The area of study was 304 km² — comprising grasslands, shrublands and the climax habitat of dry mixed evergreen forest. Rocky outcrops can be observed in patches scattered throughout the park (Jayasekara et al., 2021). Most of the grasslands and shrublands are a result of slash and burn cultivation practised over the years, until the area was declared a national park in 1983 (IUCN, 1990). The grasslands assume characteristics of savannas in some areas, whereas the reservoir perimeter is surrounded by seasonal grasses that grow during the dry season (late January-October). The park is well known for large numbers of sightings of Asian elephants (Elephas maximus) and also provides habitats for many other mammalian species (Jayasekara and Mahaulpatha, 2019) as well as avifauna (Dissanayake, 1995). The large Maduru Oya reservoir (6100 ha), constructed as a part of the Mahaweli Development Project (a large-scale national irrigation project to harness water from Sri Lanka's largest river - the Mahaweli), situated at the centre of the park, has a considerable influence on this faunal assemblage and creates a large perimeter (97.8 km) with aquatic, riparian habitats. We selected the western flank of the park for our study because the natural barriers and the man-made reservoirs/canals help in fulfilling one key assumption of both REM and CTDS models - the requirement of a closed population (Howe et al., 2017; Rowcliffe et al., 2008). Most of the study area is surrounded by four large reservoirs, irrigation canals, rock formations, and cultivated lands surround (Fig. 1) (IUCN, 1990).

Camera trapping

We conducted camera trapping mostly during the dry season (compared to the monsoon season from October to January) (IUCN, 1990) adhering to the protocol for tropical forest vertebrate camera trap survey by TEAM Network (2011). We divided the selected study area in to 2×2 km plots using a feature grid in ArcMap version 10.4.1 (ESRI, Redlands, USA) (Fig. 1). Generating this grid fulfills the spacing requirement recommended by TEAM Network (2011) of placing one camera in every 2 km² grid plot. We used two infra-red-triggered camera models: Browning Strike Force HD Pro (n=10, low glow flash) and Browning Dark OPS HD Pro (n=15, no glow flash) (Browning, USA). Except for the type of flash, the specifications of the two camera models were similar. We especially used these flash types to reduce interference to animals and meet the assumption of independent animal movement (Rowcliffe et al., 2008).

We established camera trap stations in 90 plots. We excluded plots covered with large areas of reservoir, inaccessible terrain and some plots with repetitive habitats, to obtain a balanced sampling effort in all available habitat types (Rovero et al., 2013). We deployed the moving survey method (Palencia et al., 2021b) to better use the available cameras which increase the sampling effort and precision. One station had to be excluded from analyses because a camera was stolen by a poacher, reducing the total sampling points to 89. We randomly selected plots and we placed cameras within each selected plot moving in a random distance from a random starting point in the grid line of the plot gird (walking perpendicularly to the grid line). This randomisation of camera stations fulfills the requirement of both REM and CTDS methods. Usually, we attached cameras at a fixed height of 25 cm to tree trunks or an erected log. We selected this height based on previous literature (Kalle, 2013) and our field experience of camera trapping meso-mammal carnivores, to maximise detection. We oriented cameras in a northward direction. We had to deviate the realised sampling locations and orientations up to a maximum of 100 m and 40° respectively to ensure cameras were mounted at suitable locations without obstructions (Pfeffer et al., 2018; Howe et al., 2017). However, we remained as close as possible to the predefined coordinates and orientation. We ensured mounting cameras parallel to the ground and to avoid areas with slopes, to obtain accurate distance measurements during analyses. We used protective metal cases and python lock cables when mounting cameras, to reduce damage from elephant attacks and theft. We set all cameras to function for 24 hours in a stretch of 38.2 days on average. We set the range parameter to "long range", mode of capture to "video" and trigger delay to one second. These specifications ensured that capture data could be used for both the REM and CTDS methods. We monitored the camera stations on a routine basis of 10-15 days and stations with defects in cameras/memory cards were resampled to obtain the desired sampling effort. We had to reassign two camera stations where initial coordinates coincided with resting places of a fishing cat and ring-tailed civets.

Random Encounter Model

We used REM developed by Rowcliffe et al. (2008) as one method of meso-mammal carnivore density (Dkm^{-2}) estimation. The equation

$$D = \frac{y}{t} \times \frac{\pi}{vr(2+\theta)}$$

is used for the calculation where y denotes the number of capture events; t the survey effort (camera trapping days); v the average daily distance travelled (km/day); r the average distance to the first capture of animals (km); and the average angle to the capture animals is θ (radians). The daily distance travelled (v, day range) is derived using the movement speed (s) and activity level (a) of animals following the equation shown below:

 $v = s \times a$.

The movement speed (*s*) of each animal was derived using the simple equation $s_i = \frac{d_i}{t_i}$ (Pfeffer et al., 2018) where d_i denotes the distance travelled and t_i the time duration. We followed the procedure described by Rowcliffe et al. (2016) to calculate the average speed parameter by fitting probability distributions to samples of individual speed observations obtained from video captures instead of multiple snapshots. The R package fitdistrplus (Delignette-Muller and Dutang, 2015) was used for model fitting and best fitting models were selected based on Akaike Information Criterion (AIC) values.

To determine activity level (*a*) and the proportion of the day a species is active (Rowcliffe et al., 2016), we used the R package activity (Rowcliffe, 2019; Rowcliffe et al., 2014). We converted the time stamp data of species captured on camera trap videos to radian time and analysed this in R with 1000 iterations.

To determine the radial distance (r) to the capture animal and d_i , accurate evaluation of distance from the camera was highly important. The method generally used for distance estimation is based on marking certain distance intervals from the camera at the time of mounting camera traps (Palencia et al., 2021b; Pfeffer et al., 2018; Caravaggi et al., 2016) or measuring distances of each animal manually at time of dismounting (Rowcliffe et al., 2011). However, we found that this



Figure 2 – The distance grid superimposed on camera trap capture frame to estimate distances.

method required extra time and effort in the field and that visual estimation of distances outside the marking points was difficult. In addition, in MONP where elephant activity was quite high, spending extended time in certain locations was dangerous. Therefore, we deviated from the original method of measuring distance. Rather than measuring distances on location, we incorporated the distance intervals in a pre-marked grid (Caravaggi et al., 2016) (Fig. 2), as a standard which could be superimposed on all camera trap records. This method made the determination of distances and trigonometric calculation of distances (Pfeffer et al., 2018; Caravaggi et al., 2016) easier and accurate (a distance-angle table generated following this method is given in Tab. S2). We calculated the time difference (t_i) from the time difference recorded in each video capture. Instead of camera specific detection distance and angles (Rowcliffe et al., 2008), for our analyses, we used species specific average detection distances (ADD); average detection angles (θ , ADA) derived exclusively from camera trap captures (Pfeffer et al., 2018). Because most of the observed species are solitary species we did not apply the group size function to the density equation (maximum average group size recorded was 1.06). We performed density calculations in R version 4.0.3 (R Core Team, 2013) bootstrapping with 1000 iterations from the original data.

Camera trap distance sampling method

The CTDS method, developed by Howe et al. (2017), follows the standard point transect methods (Buckland et al., 2001) and each camera station is considered a sample point. Density (D) is estimated as

$$\hat{D} = \frac{\sum\limits_{k=1}^{K} n_k}{\pi w^2 \sum\limits_{k=1}^{K} e_k \hat{p}_k}$$

where

- k = the camera station/point
- K = set of camera stations/points
- n = number of captures
- w = truncation distance beyond which any recorded distances are discarded
- e_k = effort expended at point k
- \hat{P}_k = estimated probability of obtaining an image of an animal that is within θ and w in front of the camera at a snapshot moment.

The effort is described by $e_k = \frac{\theta T_k}{2\pi t}$, that multiplied by the activity level (*a*), yields the actual trapping effort as follows:

$$e_k = \frac{\theta T_K}{2\pi t} a$$

where

- θ = average detection angle
- T_k = time period the camera was active
- t = the time between two snapshot moments considered (within the video)

Detailed explanations of these equations are provided in Howe et al. (2017). We calculated the effort for each camera trap station for separate species and provided it as the input for effort in distance software. Because of low height of the camera mount, the w values exceeding 6.2 m were less accurate. Therefore, we right truncated to a maximum of 6.2 m and left truncated to 1 m. We used the previously calculated a values (for REM) in this equation. Detection angle θ was estimated as 0.715585 radians. We recorded the distance between cameras and animals every three seconds in video captures, 24 h per day. Hence, parameter t applied in the above equation was three seconds. A special consideration was given for observations of reactivity to the cameras by the animals. In such cases, the latter part of the videos where animals unusually stayed extended time periods in front of the cameras were excluded from analysis. We used the "Distance 7.1" software package (Thomas et al., 2010) for density calculations. Half-normal and hazard rate candidate models of the detection function were tested setting the maximum adjustment parameter at one to reduce overfitting with overly complex models (Cappelle et al., 2021; Howe et al., 2019; Thomas et al., 2010; Marques et al., 2007). Fitted probability density and detection probability plots were inspected to ensure they were monotonically non-increasing (Cappelle et al., 2021; Howe et al., 2019). Competing models with sufficient goodness of fit were selected using AIC criteria.

We estimated variances in Distance 7.1 using the default analytic variance estimators based on detection probability and encounter rate (Fewster et al., 2009), and also from 1000 non-parametric bootstrap resamples of camera station data points (Cappelle et al., 2021; Howe et al., 2017; Buckland et al., 2001). Bootstrap density estimates were recorded separately. Coefficient of variation (CV) was obtained using the square root of the variance and the point estimates in all methods used.

Density estimations were compared statistically using the Wald test, with a test statistic W assessed on the chi-squared distribution with one degree of freedom (Palencia et al., 2021b; Wald and Wolfowitz, 1940). We calculated relative abundance index (RAI) as a crude estimate (Cappelle et al., 2021) for all species, especially to represent the less abundant species where sufficient samples were lacking to calculate density. RAI was calculated as encounters per hundred trap nights (Kalle, 2013).

Results

Meso-carnivore assemblage and capture abundance

A total of 3402 camera trapping days yielded 3357 video captures of 69 different animal taxa including 658 video captures of meso-mammal carnivores. During this study in MONP, we recorded all 12 mesomammal carnivore species (Tab. S1 in Supplemental Materials) found on the island. However, we captured only seven species in excess of 45 videos. The abundance of rusty spotted cats (n=4) (Fig. 3A), jungle cats (n=5), common palm civets (n=2), and brown mongooses (n=10) was very low in the study site. Therefore, density calculations based on REM model were performed only for the remaining species: fishing cats (n=106) (Fig. 3C), ruddy mongooses (n=302), stripe-necked mongooses (n=52) (Fig. 3B), ring-tailed civets (n=118) (Fig. 3D), golden palm civets (n=45) (Fig. 3E), otters (n=46) and golden jackals (n=45). We estimated that these capture numbers are greater than, or closer, to the benchmark of "around 50" captures recommended by Rovero et al. (2013) for REM density estimates. Density calculations for the same species were also conducted based on CTDS method.

Detection distances/movement speeds/day ranges, activity patterns and activity level

The average detection distance (ADD) value ranged from 1.90–4.07 m for the species considered. The rusty-spotted cat recorded the lowest distance value, while otter recorded the highest. In general, effective

detection distance (EDD) values were greater than the observed ADD values except for golden palm civet (Tab. 1).

The movement speeds ranged from 0.72–3.42 km/h. The fastest moving species was the otter, followed by the golden jackal, resulting in high day ranges for those two species. The highest activity levels were shown by fishing cat and golden jackal indicating that they were active during a greater proportion of time when compared to other species. We observed that all mongoose species and golden jackals were diurnal while civet species and otters were nocturnal (Tab. 1; Fig. 4). Fishing cats were mostly nocturnal yet could also be observed during day time as well. Jungle cats and rusty-spotted cats were recorded mostly at night. The highly nocturnal golden palm civet was the least active species. In addition, we observed this species to be the second slowest, recording the lowest day range (3.47 km/day).

Comparison of REM and CTDS density estimates

Based on Wald test statistic, any of the density estimates obtained from different methods of analyses were not significantly different for any of the species (p>0.05). However, the density estimates of fishing cat (Wald test: CTDS vs. REM: W=0.91, p=0.34) and ring-tailed civet (Wald test: CTDS (b) vs. REM: W=2.06, p=0.15) obtained using REM were relatively higher than CTDS estimates (Tab. 2). Ruddy mon-gooses had the highest abundance, and it was among the highest density estimates in all three analyses. However, the REM density estimate of ring-tailed civet was the highest recorded density. Lowest densities were recorded for otter and golden jackal. Density estimates derived using the CTDS method generally yielded lower figures when compared to the REM method (except on two occasions) (Tab. 2). However, the coefficient of variation (CV) values were generally higher in the CTDS method compared to REM (except in one occasion) (Tab. 2). The low

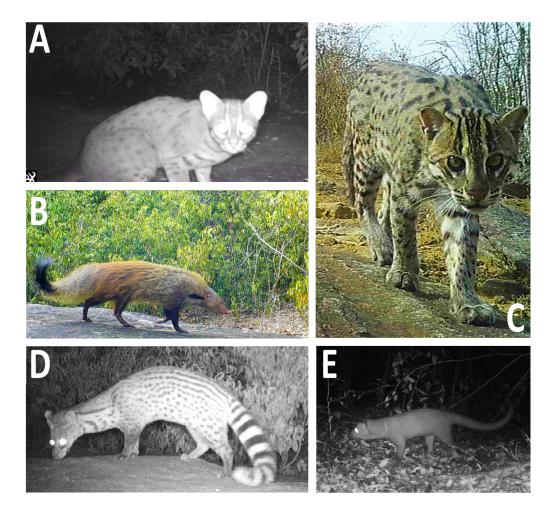


Figure 3 – (A) Rusty-spotted cat, (B) Stripe-necked mongoose (C) Fishing cat (D) Ring-tailed civet and (E) Golden palm civet captured in our camera traps.

Table 1 – Additional parameters derived for density calculations. ADD: average detection distance; EDD: effective detection distance; Speed of animal movement; Activity pattern (sD = strictly diurnal; sN = strictly nocturnal; mD = mainly diurnal; mN = mainly nocturnal; Activity level: the proportion of the day a species is active); Day range: daily distance travelled; IUCN status (IUCN, 2021).

Species	ADD (m)	EDD (m)	Movement Speed (km/h)	Activity pattern	Activity level	Day range (km/day)	IUCN status (Global)
Fishing cat Prionailurus viverrinus	2.54	2.75	0.72	mN	0.461	7.96	VU
Rusty-spotted cat Prionailurus rubiginosus	1.9	-	-	mN	-	_	NT
Jungle Cat Felis chaus	2.62	-	-	mN	-	_	LC
Ring-tailed civet Viverricula indica	2.84	3.19	1.02	sN	0.288	7.05	LC
Golden palm civet Paradoxurus zeylonensis	3.01	2.89	0.86	sN	0.161	3.34	LC
Stripe-necked mongoose Urva vitticollis	3.11	3.35	1.22	sD	0.288	8.39	LC
Ruddy mongoose Urva smithii	2.91	3.47	1.84	sD	0.390	17.22	LC
Brown mongoose Urva fuscus	2.92	-	-	sD	-	_	LC
Otter Lutra lutra	4.07	4.92	3.42	mN	0.353	28.97	NT
Golden jackal Canis aureus	3.35	3.97	3.10	mD	0.419	31.13	LC

abundance of rusty-spotted cat, jungle cat and brown mongoose were indicated by very low RAI values (Tab. 2).

Discussion

Our findings show that MONP is a protected area with a rich assemblage of meso-mammal carnivores (Tab. S1 in Supplemental Materials). However, when Felid species were considered, there were very few jungle cat and rusty-spotted cat camera trap sightings inside the study area of MONP. Because of the low number of captures of those two species, we were unable to calculate population densities using the REM or CTDS. However, RAI values of jungle cat and rusty-spotted cat were the lowest among the species on which we focused. The limited number of records of rusty spotted cats and jungle cats were from dense dry mixed evergreen forests and shrublands respectively, confirming the findings of Bora et al. (2020); Chatterjee et al. (2020) and Palei et al. (2019) on these cats' habitat occupancy. Based on our field observations, we posit tentatively that one reason for the low abundance could be that these two species are attracted to agricultural areas (paddy fields) and habitat edges, alternative habitats with abundant small mammal prey used by both species (Dharmarathne, pers. comm., 2021; SCAR, 2021; Bora et al., 2020; Miththapala, 2018; Šálek et al., 2010; Nekaris, 2003). In contrast, our results indicate that MONP is home to a healthy population of fishing cats, the largest of the three felid species studied. The fishing cat population densities recorded in this study are among the highest densities recorded for the species compared to research in other countries (Mishra et al., 2018; Sathiyaselvam et al., 2016). The large Maduru Oya reservoir and other reservoirs within the park provide ample food for this carnivore that is associated with water (SCAR, 2021; Ganguly and Adhya, 2020; Hunter, 2019; Miththapala, 2018; Mukherjee et al., 2016). The frequent release of fingerlings to the Maduru Oya reservoir by the local community-based fishing society and the abundance of fish and aquatic avifauna in its habitats make MONP an ideal site for fishing cats through the provision of food re-

			Density (individuals per km ²)			
Species	RAI	Density Estimate Method	Estimate	LCL	UCL	% CV
		REM	1.54	0.82	2.39	29.0
Fishing cat P. viverrinus	3.11	CTDS(b) (hr)	1.13	0.45	2.90	50.1
		CTDS (hr)	0.90	0.51	1.60	29.8
Rusty-spotted cat P. rubiginosus	0.12	-	-	-	-	-
Jungle cat F. chaus	0.15	-	-	-	-	-
Ring-tailed civet V. indica		REM	2.28	1.13	3.57	35.8
	3.47	CTDS(b) (hr)	1.91	1.03	3.55	31.9
		CTDS (hr)	1.69	1.09	2.63	22.6
Golden palm civet P. zeylonensis		REM	1.69	1.17	2.29	20.8
	1.32	CTDS(b) (hn)	0.80	0.32	1.98	48.6
		CTDS (hn)	0.97	0.42	2.24	44.2
Stripe-necked mongoose U. vitticollis		REM	0.75	0.47	1.06	23.0
	1.53	CTDS(b) (hr)	0.62	0.32	1.22	34.9
		CTDS (hr)	0.56	0.34	0.93	26.1
Ruddy mongoose U. smithii		REM	2.19	1.48	2.95	21.3
	8.88	CTDS(b) (hr)	2.32	1.37	3.93	27.1
		CTDS (hr)	2.23	1.40	3.56	23.9
Brown mongoose U. fuscus	0.29	-	-	-	-	-
Otter L. lutra		REM	0.15	0.05	0.28	45.9
	1.35	CTDS(b) (hr)	0.16	0.07	0.36	45.0
		CTDS (hr)	0.15	0.05	0.47	61.1
Golden jackal C. aureus		REM	0.17	0.07	0.27	39.1
	1.32	CTDS(b) (hn)	0.16	0.60	0.42	51.67
		CTDS (hn)	0.16	0.07	0.40	48.5

Table 2

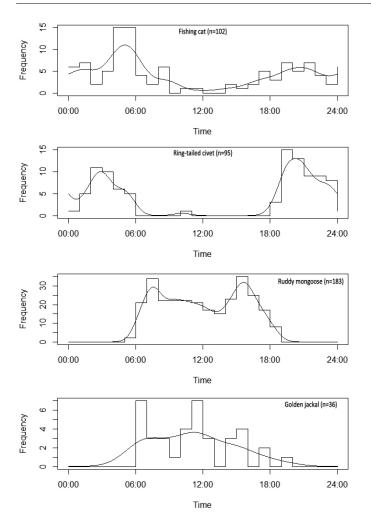


Figure 4 – Activity patterns of fishing cat, ring-tailed civet, ruddy mongoose and golden jackal in MONP, as captured by distributions of camera-trap records. Black steps are observed frequencies, and curves are fitted circular kernel distributions.

sources (Ganguly and Adhya, 2020; Hunter, 2019; Cutter, 2015; Kitchener et al., 2010; Haque and Vijayan, 1993).

Our results show that density of otters was relatively low, although this is another species that prefers aquatic fauna as its main prey (Dettori et al., 2021; Romero and Guitián, 2017; Bouros and Murariu, 2017; de Silva, 1996; Carss, 1995). Although their population density (maximum estimate 0.16 per km²) is similar to estimations from other studies (Quaglietta et al., 2015; Hájková et al., 2009; Lanszki et al., 2008), there may be a foraging niche overlap with fishing cat given their known food habits (Dettori et al., 2021; Ganguly and Adhya, 2020; Hunter, 2019; Cutter, 2015; de Silva, 1996; Kitchener et al., 2010; Carss, 1995; Haque and Vijayan, 1993).

This is likely the first effort of estimating the densities of civets and mongooses in a wild habitat in Sri Lanka. The grey mongoose, which is thought to be common in the northern third of the island (Wijeyeratne, 2008), was not captured in camera traps, although through direct visual observations, we spotted a couple of individuals. Santiapillai et al. (2000) and Wijeyeratne (2008) have reported a similar situation from Yala National Park, which is another protected area situated in the dry zone of the country. The brown mongoose abundance in MONP was low. The density of stripe-necked mongoose was moderate. We obtained high population density estimates among all focal species for the ruddy mongoose, which was also the dominant mongoose species in MONP, as observed by Jayasekara and Mahaulpatha (2019). The ring-tailed civet was the Viverrid with the highest density, validating its least concern (LC) status in the National Red List (MOE, 2012). The common palm civet density was not calculated because of the very low number of captures.

When the two main analysis methods (REM and CTDS models) are compared, the only contrasting result we obtained was the density of endemic golden palm civet. Golden palm civets are generally arboreal (Wijeyeratne, 2008) and the camera traps capture them only when they are on the ground. Therefore, the speed estimation based on a 2D model becomes biased, because their vertical movements were not recorded through our camera arrangement. The slowness of golden palm civets on ground is indicated by our speed calculation of 0.89 km/h. Considering the above, we recommend that the CTDS estimates (0.80-0.97 individuals per km²) in which speed is not a parameter, to be relatively more accurate for this species despite the drawback of not recording arboreal movements. However, the bias caused by not recording vertical movements would not be completely eliminated unless methodology is adapted to account for such complex scenarios. In general the CTDS method is considered more suitable for low abundant species (Palencia et al., 2021b).

The unusually high "speed parameters" generated for otter and golden jackal did not have an adverse impact on REM density estimation because we obtained similar densities from the CTDS method. We suggest estimating the day ranges of the above two species in the study area using another method/repeated method to confirm the values we received. However, according to Rowcliffe et al. (2012) and Palencia et al. (2019) the alternative methods such as telemetry often underestimate travel distances. Radio tracking studies of otter in other countries indicate that otters can cover long distances ranging from >20–100 km in a single day (Ruiz-Olmo et al., 2001) and occupy large home ranges (Quaglietta et al., 2015). Therefore, the day range of 27.5 km observed in the present study could likely be accurate. Research focused on golden jackal in Sri Lanka remains scarce (Jayaratne and Seneviratne, 2020) and the observed density value was within the density range observed by Šálek et al. (2014) in Balkan Peninsula.

Approximately similar density estimates generated by both analyses, despite REM estimates being slightly higher, conform the observations of Palencia et al. (2021b). Therefore, we recommend both REM and CTDS methods for the population density estimation for mesomammal carnivores in tropical habitats. However, CV values of CTDS method were relatively higher than the REM values despite the similarities of density figures. Density estimates of species with CV values <40% are generally considered reasonable, and in recent research work, the effort has been to further increase the precision (Cappelle et al., 2021; Palencia et al., 2021b; Harris et al., 2020; Howe et al., 2019). According to Cappelle et al. (2021), CV values between 10-20% are more desirable. When the present study is considered, 61.9% of the CV values were <40% and 42.9% were <30%. According to recent research, the precision can be further increased by increasing the sampling effort in different ways (Cappelle et al., 2021; Rovero et al., 2013). Hence, in order to obtain a greater number of capture events, we suggest following the recommendations of Cappelle et al. (2021); (a) increase the number of camera stations or (b) increase the length of sampling period. However, there remains the logistic concerns that are associated when camera trapping extremely rare species. We suggest the length of sampling period to be increased while deploying the appropriate number of camera stations as the best way forward. The moving survey method we followed also reduced the limitation occurred by low number of cameras, increasing the effort and precision.

Accounting for overdispersion with more customized model selection criteria as described by Howe et al. (2019) would increase the accuracy and precision of CTDS results. We identified that proper estimation of movement speed, activity and ultimately the day range of species was critical for the final density results of REM. Application of recently developed method by Palencia et al. (2019) integrating the behaviors and speed-ratio in calculations makes it possible to obtain unbiased day range values. Furthermore, with the development of machine learning techniques (Palencia et al., 2021a) and specialised R packages like trappingmotion (Palencia, 2020), the analysis process will be streamlined. However, dealing with multiple species, we observed that number of encounters need to be higher in order to apply this method. When monitoring gregarious species, it is recommended to consider applying the group size function in the density equations (Rowcliffe et al., 2008). During the present study, the ruddy mongoose and the golden jackal were the species with the highest average group size with a value closer to one (1.06). Therefore, we did not include group size in the analyses.

We used a modified distance measuring method for this study, which saved the time and effort during field work and further helped to obtain accurate measurements during analyses. However, we would like to highlight that if the distance grid and table are used, camera height and orientation should be positioned precisely. In addition, based on the camera mounting height, this distance grid and table can be generated easily prior to camera trap deployment in the field. It is also important to note that the focal distance of the camera may differ from one model to another. When using different camera models, model specific distance calculations should be used. This method is less applicable in complex field situations with slope and rugged terrain. In those instances, original distance measuring techniques or slope adjusted parameters can be used. Both REM and CTDS methods require reasonable amount of field effort as well as substantial amount of time for processing the images/videos and exploratory analyses (Palencia et al., 2021b). We would like to highlight the requirement of suitable software for image and especially video processing. Integration of such software with machine learning would greatly reduce the time required in computer analyses.

The type of camera flash also has an impact on the behaviour and the movement speed of the animals. We highly recommend a no glow flash model such as Browning Dark OPS HD Pro, which causes minimum interference to the animals when REM and CTDS methods are used. However, we observed that the low glow flash Browning Strike Force HD Pro also interfered less, except for a few observations which we had to discard the capture records as behavioural changes were observed. Selection of these flash types also increases the battery life of cameras (one set of batteries usually lasted more than two months on video mode during our study). We do not recommend white flash camera models. Most of the focal species did not react to the cameras in a greater proportion of encounters. However, there were several instances where fishing cats and ring-tailed civets were observing the cameras in an enthusiastic nature where we had to discard some parts of the videos. Though not focused on in this study, elephants were highly reactive to the cameras and were often found attacking them. The use of videos (Cappelle et al., 2021; Howe et al., 2017) - instead of snapshots used in early REM and CTDS based studies (Pfeffer et al., 2018; Rovero and Marshall, 2009) - improves the accurate identification of species. Moreover, the ability to observe the actual behaviour of the animal helps to determine when reactive behaviours take place. This also helped us to identify resting places of animals, which led to redeployment of two camera stations. Because we assessed multiple meso-carnivore species in this study, there was the concern of selecting a camera height that suits all species. Based on our observations, the increase in species shoulder height did not adversely impact the detection or encounter rate. Sometimes, the species could be identified even when some parts of the animal were out of the frame (for example the Jackal). We selected the height of 25 cm to reduce the bias caused by not encountering the smaller animals when they are very close to the camera (for example the rusty-spotted cat). Therefore, the selection of camera height should be based on the morphometrics of the focal species. The availability of in-built display with video playback option was very useful during routine observations in the field. In addition, with videos, the movement speed estimation becomes more accurate because the bias caused by the delay between the snapshots is removed. Even though the methods followed during our work would have reduced the bias of animal reactivity and other technical concerns, we acknowledge that they were not eliminated completely.

Our study provides population density estimates for the mesomammal carnivore species in MONP, which would inform future conservation and management decision-making and also a template by which their status could be assessed in forest habitats in other parts of the island. Additional parameters such as movement speed, activity patterns, activity level and day range that we generated can be also used for future research in a broad range of applications. The study shows clearly that REM and CTDS methods can be applied practically under field conditions of tropical forests, to assess multiple species. The recommendations for modifications to build upon original methodologies and analyses will improve efficiency and cost-effectiveness of similar research in the future.

Conclusions

The study identifies MONP as a protected area with a rich mesomammal assemblage. However, our study indicates that species such as rusty-spotted cat, jungle cat, brown mongoose, otter and golden jackal have low abundances and population densities. MONP sustains considerably healthy populations of fishing cats, ring-tailed civets and ruddy mongooses. The two main population estimation methods we used, the REM method and CTDS method could be applied successfully in the forest habitats of Maduru Oya. The CTDS method was more easily applicable in the field with suggested modifications of distance estimations. However, the relatively complex REM method can be more useful as it generates additional information such as activity, day range and movement speed which are useful for other ecological studies and decision making.

References

- Anile S., Ragni B., Randi E., Mattucci F., Rovero F., 2014. Wildcat population density on the Etna volcano, Italy: a comparison of density estimation methods. J. Zool. 293(4): 252–261.
- Bater C.W., Coops N.C., Wulder M.A., Hilker T., Nielsen S.E., McDermid G., Stenhouse G.B., 2011. Using digital time-lapse cameras to monitor species-specific understorey and overstorey phenology in support of wildlife habitat assessment. Environ. Monit. Assess. 180(1): 1–13.
- Bessone M., Kühl H.S., Hohmann G., Herbinger I., N'Goran K.P., Asanzi P., Da Costa P.B., Dérozier V., Fotsing E.D., Beka B.I., Iyomi M.D., 2020. Drawn out of the shadows: Surveying secretive forest species with camera trap distance sampling. J Appl Ecol. 57(5): 963–974.
- Bora J.K., Awasthi N., Kumar U., Goswami S., Pradhan A., Prasad A., Laha D.R., Shukla R., Shukla S.K., Qureshi Q., Jhala Y.V., 2020. Assessing the habitat use, suitability and activity pattern of the Rusty-spotted Cat *Prionailurus rubiginosus* in Kanha Tiger Reserve, India. Mammalia 84(5): 459–468.
- Bouros G., Murariu D., 2017. Comparative diet analysis of the Eurasian otter (*Lutra lutra*) in different habitats: Putna-Vrancea Natural Park and Lower Siret Valley, south-eastern Romania. North West J Zool. 13(2): 311–319.
- Bowkett A.E., Rovero F., Marshall A.R., 2008. The use of camera-trap data to model habitat use by antelope species in the Udzungwa Mountain forests, Tanzania. Afr J Ecol. 46(4): 479–487.
- Buckland S.T., Anderson D.R., Burnham K.P., Laake J.L., Borchers D.L., Thomas L., 2001. Introduction to Distance Sampling. Estimating Abundance of Biological Populations. Oxford University Press, Oxford, UK.
- Buckland S.T., Anderson D.R., Burnham K.P., Laake J.L., Borchers D.L., Thomas L., 2004. Advanced Distance Sampling. Estimating Abundance of Biological Populations, Oxford University Press, Oxford, UK.
- Buckland S.T., Rexstad E.A., Marques T.A., Oedekoven C.S., 2015. Distance Sampling. Methods and Applications, Springer, Heidelberg, Germany.
- Cappelle N., Després-Einspenner M., Howe E.J., Boesch C., Kühl H.S., 2019. Validating camera trap distance sampling for chimpanzees. Am. J. Primatol. 81(3): e22962. https://doi.org/10.1002/ajp.22962
- Cappelle N., Howe E.J., Boesch C., Kühl H.S., 2021. Estimating animal abundance and effort-precision relationship with camera trap distance sampling. Ecosphere 12(1): p.e03299.
- Caravaggi A., Zaccaroni M., Riga F., Schai-Braun S.C., Dick J.T., Montgomery W.I., Reid N., 2016. An invasive-native mammalian species replacement process captured by camera trap survey random encounter models. Remote Sens. Ecol. Con. 2(1): 45–58.
- Carbone C., Christie S., Conforti K., Coulson T., Franklin N., Ginsberg J.R., Griffiths M., Holden J., Kawanishi K., Kinnaird M., Laidlaw R., 2001. The use of photographic rates to estimate densities of tigers and other cryptic mammals. Anim. Conserv. 4(1): 75–79.
- Carss D., 1995. Foraging behaviour and feeding ecology of the otter *Lutra lutra*: a selective review. Hystrix 7: 179–194.
- Chandler R.B., Royle J.A., 2013. Spatially Explicit Models for Inference About Density in Unmarked or Partially Marked Populations. Ann. Appl. Stat. 7(2): 936–954.
- Chatterjee N., Nigam P., Habib B., 2020. Population density and habitat use of two sympatric small cats in a central Indian reserve. PLoS ONE 15(6): p.e0233569.
- Clevenger A.P., Ford A.T., Sawaya M.A., 2009. Banff wildlife crossings project: Integrating science and education in restoring population connectivity across transportation corridors. Final report to Parks Canada Agency, Radium Hot Springs, British Columbia, Canada. Available from http://arc-solutions.org/wp-content/uploads/2012/03/ Clevengeret-al-2009-Ban_-wildlife-crossings-project.pdf [Accessed 3 February 2020]
- Corlatti L., Sivieri S., Sudolska B., Giacomelli S., Pedrotti L., 2020. A field test of unconventional camera trap distance sampling to estimate abundance of marmot populations. Wildlife Biol. 2020(4): 1–11. 10.2981/wlb.00652
- Cusack J.J., Swanson A., Coulson T., Packer C., Carbone C., Dickman A.J., Kosmala M., Lintott C., Rowcliffe J.M., 2015. Applying a random encounter model to estimate lion density from camera traps in Serengeti National Park, Tanzania. J. Wildl. Manag. 79(6): 1014–1021.

- Cutter P., 2015. Fishing cat ecology: food habits, home ranges, habitat use and mortality in a human-dominated landscape around Khao Sam Roi Yot, Peninsular Thailand. M. Sc. thesis. Available from the University of Minnesota Digital Conservancy. https://hdl. handle.net/Il299/I74755
- de Silva P.K., 1996. Food and feeding habits of the Eurasian Otter Lutra lutra L. (Carnivora, Mustelidae) in Sri Lanka. J. South Asian Nat. Hist. 2(1): 81–90.
- Dettori E.E., Balestrieri A., Zapata-Perez V.M., Bruno D., Rubio-Saura N., Robledano-Aymerich F., 2021. Distribution and diet of recovering Eurasian otter (*Lutra lutra*) along the natural-to-urban habitat gradient (river Segura, SE Spain). Urban Ecosyst. 2021: 1– 10. 10.1007/s11252-021-01109-3
- Delignette-Muller M.L., Dutang C., 2015. fitdistrplus: An R Package for Fitting Distributions. J. Stat. Softw. 64(4): 1–34. https://www.jstatsoft.org/v64/i04/
- Dissanayake S.R.B., 1995. Avifaunal ecology with respect to different habitat types in the Maduruoya National Park. M. Phil. thesis, Division of Zoology, Faculty of Natural Sciences, The Open University of Sri Lanka, Nawala, Sri Lanka.
- Dittus W.P., 2017. The biogeography and ecology of Sri Lankan mammals point to conservation priorities. Ceylon J. Sci. 46(Special Issue): 33–64.
- Engeman R.M., Massei G., Sage M., Gentle M.N., 2013. Monitoring wild pig populations: a review of methods. Environ. Sci. Pollut. Res. 20(11): 8077–8091.
- Fewster R., Buckland S., Burnham K., Borchers D., Jupp P., Laake J., Thomas L., 2009. Estimating the encounter rate variance in distance sampling. Biometrics 65: 225–236.
- Estimating the encounter rate variance in distance sampling. Biometrics 65: 225–236. Ganguly D., Adhya T., 2020. How Fishing Cats *Prionailurus viverrinus* fish: Describing a felid's strategy to hunt aquatic prey. bioRxiv. 10.1101/2020.04.24.058925
- Gilbert N.A., Clare J.D.J., Stenglein J.L., Zuckerberg B., 2020. Abundance estimation of unmarked animals based on camera-trap data. Conserv. Biol. 35(1): 88–100.
- Glover-Kapfer P., Soto-Navarro C.A., Wearn O.R., 2019. Camera-trapping version 3.0: current constraints and future priorities for development. Remote Sens. Ecol. Con. 5(3): 209–223.
- Gray T.N.E., 2018. Monitoring tropical forest ungulates using camera-trap data. J. Zool., 305(3): 173–179.
- Green S.E., Rees J.P., Stephens P.A., Hill R.A., Giordano A.J., 2020. Innovations in camera trapping technology and approaches: The integration of citizen science and artificial intelligence. Animals 10(1): 132.
 Hájková P., Zemanová B., Roche K., Hájek B., 2009. An evaluation of field and noninvasive
- Hájková P., Zemanová B., Roche K., Hájek B., 2009. An evaluation of field and noninvasive genetic methods for estimating Eurasian otter population size. Conserv. Genet. 10(6): 1667–1681.
- Haque N.M., Vijayan V., 1993. Food habits of the fishing cat *Felis viverrina* in Keoladeo National Park, Bharatpur, Rajasthan. J. Bombay Nat. Hist. Soc. 90(1993): 498–500. Hardouin M., Searle C.E., Strampelli P., Smit J., Dickman A., Lobora A.L., Rowcliffe
- Hardouin M., Searle C.E., Strampelli P., Smit J., Dickman A., Lobora A.L., Rowcliffe J.M., 2021. Density responses of lesser-studied carnivores to habitat and management strategies in southern Tanzania's Ruaha-Rungwa landscape. PLoS ONE 16(3): p.e0242293.
- Harris G.M., Butler M.J., Stewart D.R., Rominger E.M., Ruhl C.Q., 2020. Accurate population estimation of Caprinae using camera traps and distance sampling. Scientific Sci. Rep. 10(1): 17729. 10.1038/s41598-020-73893-5
 Hoffmann A., Decher J., Rovero F., Schaer J., Voigt C., Wibbelt G., 2010. Field methods
- Hoffmann A., Decher J., Rovero F., Schaer J., Voigt C., Wibbelt G., 2010. Field methods and techniques for monitoring mammals. In: Eymann J., Degreef J., Hauser C., Monj, J.C., Samyn Y., Van den Spiegel D. (Eds.) Manual on field recording techniques and protocols for all taxa biodiversity inventories and monitoring. Pensoft Publishers, Sofia, Bulgaria. 482–529.
- Hofmeester T.R., Rowcliffe J.M., Jansen P.A., 2017. A simple method for estimating the effective detection distance of camera traps. Remote Sens. Ecol. Con. 3(2): 81–89.Howe E.J., Buckland S.T., Després-Einspenner M.-L., Kühl H.S., 2017. Distance sampling
- Howe E.J., Buckland S.T., Després-Einspenner M.-L., Kühl H.S., 2017. Distance sampling with camera traps. Methods Ecol. Evol. 8(11): 1558–1565. 10.1111/2041-210X.12790
- Howe E.J., Buckland S.T., Després-Einspenner M., Kühl H.S., 2019. Model selection with overdispersed distance sampling data. Methods Ecol. Evol. 10(1): 38–47. 10.1111/2041-210X.13082
- Hunter L., 2019. Carnivores of the world. 2nd ed. Princeton University Press, Princeton, New Jersey.
- IUCN (International Union for Conservation of Nature), 1990. IUCN Directory of South Asian Protected Areas. IUCN, Gland, Switzerland and Cambridge, UK. xxiv + 294. IUCN (International Union for Conservation of Nature), 2021. The IUCN red list of
- threatened species. Version 2021-1. https://www.iucnredlist.org
- Jayaratne C., Seneviratne S., 2020. Sri Lankan Jackal: The Island's True Wild Canid. Loris 29(2): 19–23.
- Jayasekara D., Kumara P.K.P.M.P., Mahaulpatha W.A.D., 2021. Mapping the vegetation cover and habitat categorization of Maduru Oya and Horton Plains National Parks using Landsat 8 (OLI) imagery to assist the ecological studies. Wildlanka 9(1): 122–135.
- Jayasekara E.G.D.P., Mahaulpatha W.A.D., 2019. Abundance and distribution of Family Herpestidae in three Protected Areas representing three Bioclimatic Zones in Sri Lanka. Proceedings of the 6th International Conference on Multidisciplinary Approaches (iCMA) 2019. Available at SSRN: https://srn.com/abstract=3497273
- Jiménez J., Nuñez-Arjona J.C., Rueda C., González L.M., García-Domínguez F., Muñoz-Igualada J., López-Bao J.V., 2017. Estimating carnivore community structures. Sci. Rep. 7(1): 1–10.
- Johansson, Ö., Samelius, G., Wikberg, E., Chapron, G., Mishra, C., Low, M., 2020. Identification errors in camera-trap studies result in systematic population overestimation. Sci. Rep. 10(1): 1–10.
- Kalle R., 2013. Ecology of sympatric small carnivores in Mudumalai Tiger Reserve, Tamil Nadu. Ph D. thesis, Saurashtra University, Rajkot, India. Available online at http://hdl. handle.net/10603/15963 [Accessed 18 June 2018]
- Kalle R., Ramesh T., Sankar K., Qureshi Q., 2013. Observations of sympatric small carnivores in Mudumalai Tiger Reserve, Western Ghats, India. Small Carniv. Conserv. 49: 53–59.
- Karanth K.U., Nichols J.D., Kumar N.S., Hines J.E., 2006. Assessing tiger population dynamics using photographic capture–recapture sampling. Ecology 87(11): 2925–2937. Kitchener A.C., Van Valkenburgh B., Yamaguchi N., 2010. Felid form and function. In:
- Kitchener A.C., Van Valkenburgh B., Yamaguchi N., 2010. Felid form and function. In: MacDonald D.W., Loveridge A.J (Eds.) Biology and Conservation of Wild Felids. Oxford University Press, Oxford. 83–106.
- Kittle A.M., Watson A.C., 2018. Density of leopards (*Panthera pardus kotiya*) in Horton Plains National Park in the Central Highlands of Sri Lanka. Mammalia 82(2): 183–187.
- Kittle A.M., Watson A.C., Fernando T.S.P., 2017. The ecology and behaviour of a protected area Sri Lankan leopard (*Panthera pardus kotiya*) population. Trop. Ecol. 58(1): 71–86.

- Lanszki J., Hidas A., Szentes K., Révay T., Lehoczky I., Weiss S., 2008. Relative spraint density and genetic structure of otter (*Lutra lutra*) along the Drava River in Hungary. Mamm. Biol. 73(1): 40–47.
- Luo G., Wei W., Dai Q., Ran J., 2020. Density Estimation of Unmarked Populations Using Camera Traps in Heterogeneous Space. Wildl. Soc. Bull. 44(1): 173–181.
- Manzo E., Bartolommei P., Rowcliffe J.M., Cozzolino R., 2012. Estimation of population density of European pine marten in central Italy using camera trapping. Acta Theriol. 57(2): 165–172.
- Marques T.A., Thomas L., Fancy S.G., Buckland S.T., 2007. Improving estimates of bird density using multiple-covariate distance sampling. The Auk 124(4): 1229–1243.
- Marques T.A., Thomas L., Martin S.W., Mellinger D.K., Ward J.A., Moretti D.J., Harris D., Tyack P.L., 2013. Estimating animal population density using passive acoustics. Biol. Rev. 88(2): 287–309.
- Meek P.D., Ballard G., Falzon G., Williamson J., Milne H., Farrell R., Stover J., Mather-Zardain A.T., Bishop J.C., Cheung E.K.W., Lawson C.K., 2020. Camera trapping technology and related advances: Into the new millennium. Aust. J. Zool. 40(3): 392–403.
- Meek P.D., Fleming P., Ballard G., Banks P., Claridge A.W., Sanderson J., Swann D. (Eds) 2014. Camera trapping: wildlife management and research (Vol. 2). CSIRO Publishing, Melbourne, Australia.
- Meek P.D., Zewe F., Falzon G., 2012. Temporal activity patterns of the swamp rat (*Rattus lutreolus*) and other rodents in north-eastern New South Wales, Australia. Aust. Mammal. 34(2): 223–233.
- Mishra R., Basnet K., Amin R., Lamichhane B.R., 2018. Fishing Cat Prionailurus viverrinus Bennett, 1833 (Carnivora: Felidae) distribution and habitat characteristics in Chitwan National Park, Nepal. J. Threat. Taxa. 10(11): 12451–12458.
- Miththapala S., 2018. An overview of the wild cats of Sri Lanka Part II. Loris 28(3): 7–47. MOE (Ministry of Environment), 2012. The National Red List 2012 of Sri Lanka. Conser-
- MOE (Ministry of Environment), 2012. The National Red List 2012 of Sri Lanka. Conservation Status of the Fauna and Flora. Ministry of Environment, Colombo, Sri Lanka. viii + 476.
- Moeller A.K., Lukacs P.M., Horne J.S., 2018. Three novel methods to estimate abundance of unmarked animals using remote cameras. Ecosphere 9(8): e02331.
- MoMD&E (Biodiversity Secretariat, Ministry of Mahaweli Development and Environment), 2019. Biodiversity Profile – Sri Lanka, Sixth National Report to the Convention on Biological Diversity, Biodiversity Secretariat, Ministry of Mahaweli Development and Environment, Sri Lanka.
- Morrison M.L., 2013. Wildlife restoration: techniques for habitat analysis and animal monitoring (Vol. 1). Island Press, Washington, DC.
- Mukherjee S., Appel A., Duckworth J.W., Sanderson J., Dahal S., Willcox D.H.A., Herranz Muñoz V., Malla G., Ratnayaka A., Kantimahanti M., Thudugala A., 2016. *Prionailurus* viverrinus. The IUCN Red List of Threatened Species 2016: eT18150A50662615.
- Nakashima Y., Fukasawa K., Samejima H., 2018. Estimating animal density without individual recognition using information derivable exclusively from camera traps. J Appl Ecol. 55(2): 735–744.
- Nekaris K.A.I. 2003. Distribution and behaviour of three small wild cats in Sri Lanka. Cat News 38(1): 30–32.
- O'Brien T.G., 2011. Abundance, density and relative abundance: a conceptual framework. In: O'Connell A.F., Nichols J.D., Karanth K.U. (Eds.) Camera traps in animal ecology. Springer, Tokyo. 71–96.
- Pal R., Bhattacharya T., Qureshi Q., Buckland S. T., Sathyakumar S., 2021. Using distance sampling with camera traps to estimate the density of group-living and solitary mountain ungulates. Oryx 55(5): 668–676. 10.1017/S003060532000071X
- Palei H.S., Palei N.C., Rath B.P., Mishra A.K., 2019. Records of the globally threatened Rusty-spotted Cat in Odisha, India. Nat. Conserv. Res. 4(3): 112–116.
- Palencia P., 2020. trappingmotion: Integrate camera-trapping in movement and behavioural studies. R package version 0.1.1. https://github.com/PabloPalencia/trappingmotion
- Palencia P., Fernández-López J., Vicente J., Acevedo P., 2021a. Innovations in movement and behavioural ecology from camera traps: day range as model parameter. Methods Ecol. Evol. 12(7): 1201–1212. 10.1111/2041-210X.13609
- Palencia P., Rowcliffe J.M., Vicente J., Acevedo P., 2021b. Assessing the camera trap methodologies used to estimate density of unmarked populations. J. Appl. Ecol. 58(8): 1583– 1592. 10.1111/1365-2664.13913
- Palencia P., Vicente J., Barroso P., Barasona J.Á., Soriguer R.C., Acevedo P., 2019. Estimating day range from camera-trap data: the animals' behaviour as a key parameter. J. Zool. 309(3): 182–190.
- Parker I., Lopez R., Silvy N., Davis D., Cathey J., 2012. Alternative methodology for handling and marking meso-mammals for short-term research. Wildl. Biol. Pract. 8(2): 20– 25.
- Pfeffer S.E., Spitzer R., Allen A.M., Hofmeester T.R., Ericsson G., Widemo F., Singh N.J., Cromsigt J.P., 2018. Pictures or pellets? Comparing camera trapping and dung counts as methods for estimating population densities of ungulates. Remote Sens. Ecol. Con. 4(2): 173–183.
- Punyawardena B.V.R., 2020. Climate. In: Mapa R. (Ed). The Soils of Sri Lanka. World Soils Book Series, Springer, Cham. 13–22. 10.1007/978-3-030-44144-9_2
- Quaglietta L., Hájková P., Mira A., Boitani L., 2015. Eurasian otter (*Lutra lutra*) density estimate based on radio tracking and other data sources. Mammal Res. 60(2): 127–137.
 R Core Team 2013 R: A language and environment for statistical computing. R Foundation
- R Core Team, 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. http://www.r-project.org
 Rademaker M., Meijaard E., Semiadi G., Blokland S., Neilson E.W., Rode-Margono E.J., 2016. First ecological study of the Bawean warty pig (*Sus blouchi*), one of the rarest
- pigs on earth. PLoS ONE, 11(4): e0151732. Ramsey D.S., Caley P.A., Robley A., 2015. Estimating population density from presence-
- absence data using a spatially explicit model. J. Wildl. Manag. 79(3): 491–499. Rich L.N., Miller D.A., Muñoz D.J., Robinson H.S., McNutt J.W., Kelly M.J., 2019. Sampling design and analytical advances allow for simultaneous density estimation of
- seven sympatric carnivore species from camera trap data. Biol. Conserv. 233: 12–20. Roemer G.W., Gompper M.E., Van Valkenburgh B., 2009. The ecological role of the mam-
- malian mesocarnivore. BioScience 59(2): 165–173. Romairone J., Jiménez J., Luque-Larena J.J., Mougeot F., 2018. Spatial capture-recapture
- design and modelling for the study of small mammals. PLoS ONE 13(6): e0198766. Romero R., Guitián J., 2017. Food and feeding habits of Eurasian otter, *Lutra lutra*, and
- American mink, *Neovison vison*, in an Atlantic island of northwest Spain. J. Vertebr. Biol. 66(2): 117–125.
- Rovero F., De Luca D.W., 2007. Checklist of mammals of the Udzungwa Mountains of Tanzania. Mammalia 71: 47–55.

- Rovero F., Marshall A.R., 2009. Camera trapping photographic rate as an index of density in forest ungulates. J. Appl. Ecol. 46(5): 1011–1017.Rovero F., Zimmermann F., Berzi D., Meek P., 2013. "Which camera trap type and how
- Rovero F., Zimmermann F., Berzi D., Meek P., 2013. "Which camera trap type and how many do I need?" A review of camera features and study designs for a range of wildlife research applications. Hystrix 24(2): 148–156.
- Rowcliffe J.M., 2019. activity: Animal activity statistics. R package version 1.3. https: //CRAN.R-project.org/package=activity
- Rowcliffe J.M., Carbone C., Jansen P.A., Kays R., Kranstauber B., 2011. Quantifying the sensitivity of camera traps: an adapted distance sampling approach. Methods Ecol. Evol. 2(5): 464–476.
- Rowcliffe J.M., Carbone C., Kays R., Kranstauber B., Jansen P.A., 2012. Bias in estimating animal travel distance: the effect of sampling frequency. Methods Ecol. Evol. 3(4): 653– 662.
- Rowcliffe J.M., Field J., Turvey S.T., Carbone C., 2008. Estimating animal density using camera traps without the need for individual recognition. J. Appl. Ecol. 45(4): 1228– 1236.
- Rowcliffe J.M., Jansen P.A., Kays R., Kranstauber B., Carbone C., 2016. Wildlife speed cameras: measuring animal travel speed and day range using camera traps. Remote Remote Sens. Ecol. Con. 2(2): 84–94.
- Rowcliffe J.M., Kays R., Kranstauber B., Carbone C., Jansen P.A., 2014. Quantifying levels
- of animal activity using camera trap data. Methods Ecol. Evol. 5(11): 1170–1179. Royle J.A., Nichols J.D., 2003. Estimating abundance from repeated presence-absence data or point counts. Ecology (84): 777–790.
- Royle J.A., 2004. N-mixture models for estimating population size from spatially replicated counts. Biometrics 60(1): 108–115.
- Royle J.A., Chandler R.B., Gazenski K.D., Graves T.A., 2013. Spatial capture-recapture models for jointly estimating population density and landscape connectivity. Ecology 94(2): 287–294.
- Ruiz-Olmo J., Saavedra D., Jiménez J., 2001. Testing the surveys and visual and track censuses of Eurasian otters (*Lutra lutra*). J. Zool. 253(3): 359–369.
- Šálek M., Červinka J., Banea O.C., Krofel M., Ćirović D., Selanec I., Penezić A., Grill S., Riegert J., 2014. Population densities and habitat use of the golden jackal (*Canis aureus*) in farmlands across the Balkan Peninsula. Eur. J. Wildl. Res. 60(2): 193–200.
- Šálek M., Kreisinger J., Sedláček F., Albrecht T., 2010. Do prey densities determine preferences of mammalian predators for habitat edges in an agricultural landscape?. Landsc. Urban Plan. 98(2): 86–91.
- Sanderson J.G., Trolle M., 2005. Monitoring elusive mammals: unattended cameras reveal secrets of some of the world's wildest places. Am. Sci. 93(2): 148–155.
- Santiapillai C., De Silva M., Dissanayake S.R.B., 2000. The status of mongooses (Family: Herpestidae) in Ruhuna National Park, Sri Lanka. J. Bombay Nat. Hist. Soc. 97(2): 208– 214.
- Sathiyaselvam P., Satyanarayana E., Kathula T., 2016. Current status and threats to fishing cat *Prionailurus viverrinus* (Bennett, 1833) in Godavari Mangroves, Andhra Pradesh, India. Indian For. 142(10): 950–832.
- SCAR (Small Cat Advocacy and Research) , 2021. SCAR web site. Available at: https: //scar.lk [Accessed 12 March 2021]
- Sheftel B.I., 2018. Methods for estimating the abundance of small mammals. Russ J Ecol. 3(3): 1–21.

- Silveira L., Jacomo A.T., Diniz-Filho J.A.F., 2003. Camera trap, line transect census and track surveys: a comparative evaluation. Biol. Conserv. 114(3): 351–355.
 Srbek-Araujo A.C., Chiarello A.G., 2005. Is camera-trapping an efficient method for sur-
- Srbek-Araujo A.C., Chiarello A.G., 2005. Is camera-trapping an efficient method for surveying mammals in Neotropical forests? A case study in south-eastern Brazil. J. Trop. Ecol. 21(1): 121–125.
- TEAM Network., 2011. Terrestrial Vertebrate Protocol Implementation Manual, v. 3.1. Tropical Ecology, Assessment and Monitoring Network, Center for Applied Biodiversity Science, Conservation International, Arlington, VA, USA.
- Thomas L., Buckland S.T., Rexstad E.A., Laake J.L., Strindberg S., Hedley S.L., Bishop J.R., Marques T.A., Burnham K.P., 2010. Distance software: design and analysis of distance sampling surveys for estimating population size. J. Appl. Ecol. 47(1): 5–14.
- Tobler M.W., Carrillo-Percastegui S.E., Powell G., 2009. Habitat use, activity patterns and use of mineral licks by five species of ungulate in south-eastern Peru. J. Trop. Ecol. 25(3): 261–270.
- Wald A., Wolfowitz J., 1940. On a test whether two samples are from the same population. Ann. Math. Stat. 11(2): 147–162. http://www.jstor.org/stable/2235872
- Webb E.L., Choo Y.R., Kudavidanage E.P., Amarasinghe T.R., Bandara U.G.S.I., Wanninayaka W.A.C.L., Ravindrakumar P., Nimalrathna T.S., Liang S.H., Chua M.A.H., 2020. Leopard activity patterns in a small montane protected area highlight the need for integrated, collaborative landscape conservation. Glob. Ecol. Conserv. 23: e01182.
- Weerakoon D.K., Goonatilake W.D.A., 2006. Taxonomic status of the mammals of Sri Lanka. Fauna of Sri Lanka: Status of Taxonomy, Research and Conservation, IUCN Sri Lanka, Colombo. 216–231.
- Weerakoon D.K., 2012. The taxonomy and conservation status of mammals in Sri Lanka. In: Weerakoon D.K., Wijesundara S. (Eds) The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora. Ministry of Environment, Colombo, Sri Lanka. 134–144.
- Wijesinghe M.R., 2006. Ecological traits of endemic small mammals in rainforests of Sri Lanka, and their implications for conservation. In: Bambaradeniya C.N.B. (Ed.) The Fauna of Sri Lanka: Status of Taxonomy, Research, and Conservation, IUCN, Colombo, Sri Lanka. 232–234.
- Wijeyeratne G.D., 2008. A photographic Guide to Mammals of Sri Lanka. New UK: Holland Publishers.
- Zimmermann F., Molinari-Jobin A., Ryser A., Breitenmoser-Würsten Ch., Pesenti E., Breitenmoser U., 2011. Status and distribution of the lynx (*Lynx lynx*) in the Swiss Alps 2005–2009. Acta Biol. Slov. 54: 74–84.

Associate Editor: F. Rovero

Supplemental information

Additional Supplemental Information may be found in the online version of this article:

 Table SI
 Checklist of meso-mammal carnivores in Sri Lanka.

 Table S2
 Distance – Angle Chart.